Involvement of Sphingosine Kinases/Sphingosine-1-Phosphate (S1P)/S1P Receptors in Breast Cancer Subtypes

Abstract

There is emerging evidence suggesting sphingolipids as critical regulators of cancer development and progression. Sphingolipids are potent bioactive lipids involved in fundamental biological processes including cell proliferation, apoptosis, angiogenesis, senescence, stress response and transformation. Ceramide, sphingosine and Sphingosine-1-phosphate (S1P) are inter-convertible sphingolipids with opposing effects on cell fate. Furthermore, S1P either acts directly on intracellular targets or through G-protein coupled S1PRs (S1P1-5) to mediate their specific effects. This review will discuss the roles of key sphingolipids, sphingosine kinases (SphKs) and S1P receptors (S1PRs) in tumor growth and acquisition of resistance to chemotherapy in four subtypes of breast cancer that are categorized based on the status of hormone receptors and human epidermal growth factor 2 (HER2) receptors.

Introduction

Breast cancer is the most frequently diagnosed cancer and is second leading cause of death among U.S. women with an estimated 232,340 new cases and 39,620 deaths in 2013 [1]. Despite significant progress in clinical efficacy, approximately 30% of patients with breast cancer will develop incurable metastatic breast disease and a promising cure for this devastating disease has yet to be discovered [2]. Current prognosis and therapy largely rely on the biological subtypes determined by the expressions of estrogen, progesterone and human epidermal growth factor 2 (HER2) receptors.

Recent studies unveiled the involvement of sphingolipid signaling in breast cancers. Sphingolipids are a family of lipids having a sphingoid backbone that mainly resides in the cell membrane to provide structural support but is also known to mediate signaling cascades involved in cell proliferation, angiogenesis, apoptosis, senescence, stress response and transformation [3-6]. The key sphingolipids are S1P and its two precursors, ceramide and sphingosine. These sphingolipids are inter-convertible within cells and their balance is tightly modulated by actions of enzymes including cell proliferation, apoptosis, angiogenesis, senescence, stress response and transformation [3-6]. The key sphingolipids are S1P and its two precursors, ceramide and sphingosine. These sphingolipids are inter-convertible within cells and their balance is tightly modulated by actions of enzymes including ceramidases, ceramide synthases, SphKs, and S1P phosphatase.

Abstract

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Breast Cancer Subtypes

Breast cancer prognosis and treatment decisions are heavily dependent on the status of hormone receptors and HER2. Although not all tumors within each subtype share the same characteristics, receptor status can be used as a guide to classify breast cancers into four biological subtypes [12-15]. The first subtype is Luminal A and it is characterized by having estrogen receptor (ER) and/or progesterone receptor (PR) while lacking HER2, and it accounts for approximately 40% of all breast cancers. Second subtype that makes up 20% of all breast cancer is Luminal B. Luminal B is characterized by weak to moderate expression of ER and/or PR and HER2 overexpression [14]. The third subtype representing 10-20% of breast cancer lacks all three receptors and is referred to as triple negative/basal like tumors. The last subtype, HER2 type, overexpresses HER2 but lacks hormone receptors and this group accounts for 15-20% of breast cancers.

Estrogen and progesterone help regulate early mitogenesis and development in the mammary gland, and they are necessary for cyclic proliferation during menstrual cycle and for lobulo-alveolar growth in mammary tissue during pregnancy [16]. However, increased expressions of either estrogen alone or both estrogen and progesterone increase breast cancer risk. ERs are expressed in approximately 70% of diagnosed breast cancers and selective modulators (SERMs), such as tamoxifen and fulvestrant, are the first-line therapies for these cancers [17]. Treatment of ER-positive breast cancer patients with tamoxifen reduce disease 5-year recurrence by 50%, however, patient mortality remains high [18]. Mechanistically, ER in the nucleus functions as a transcriptional regulator and binding of Estradiol leads to a conformational change and receptor dimerization [19]. The ligand/receptor complex binds directly or indirectly to estrogen response elements in the promoter regions of estrogen-responsive gene enhancing transcription, including PR [19,20]. In general, patients with tumors expressing both ER and PR respond better to endocrine therapy and have longer survival times and later on set of recurrence compared with those tumors lacking both receptors [18,21-23].

The HER2 gene, also known as neu/c-erbB2, encodes a 185-kDa transmembrane receptor tyrosine kinase. It is a member of the epidermal growth factor (EGF) receptor (EGFR) family and it is overexpressed in approximately 30% of primary breast cancers.
Patients with tumors overexpressing HER2 have increased tumor invasion, poor prognosis, and therapeutic resistance [24]. HER2 is more potent oncoprotein compared to other EGFRs and acts independently of ligands unlike other members in the EGFR family. Instead, it acts by forming homodimers or heterodimers with other EGFRs [26]. This results in slower endocytosis and rapid recycling of EGFRs and prolonged stimulation of the extracellular signal regulated kinase (ERK) pathway, which leads to increased cell proliferation, migration and resistance to apoptosis [26,27]. Patients with tumors overexpressing HER2 respond better to immunotherapy such as anti-HER2 antibody trastuzumab (Herceptin) than those with basal HER2 levels.

Triple negative/basal like tumor subtype is defined by the absence of all three (ER/PR/HER2) receptors. Lack of receptors limits the use of advanced treatments such as endocrine-and immune-therapies, leading to a high proportion of disease-related death compared to other subtypes [28].

Role in Cancer: Sphingolipid Metabolism and Signaling

Accumulating evidence suggests that sphingolipid pathway is involved in carcinogenesis, such as colon, prostate, and head and neck as well as breast carcinogenesis [6,29]. Particularly, great emphasis has been placed on ceramide, sphingosine and S1P. The central component of the sphingolipid pathway is the ceramide, which can either be synthesized de novo from serine and palmitate or generated by the breakdown of sphingomyelin [7]. Ceramide can be hydrolyzed by actions of many ceramidases into sphingosine, which in turn is phosphorylated by SphK to form S1P. S1P can either act via S1PRs at membrane surface to regulate downstream signaling or be degraded irreversibly in the endoplasmic reticulum by S1P lyase [11,30,31]. Ceramide, sphingosine and S1P are readily inter-convertible and the balance between these sphingolipids is tightly regulated by SphKs, S1P phosphatase and type 2 phosphatidate phosphohydrolase. These three major sphingolipid metabolites have been the focus of many studies as they possess opposing biological functions where ceramide and sphingosine regulate stress responses such as apoptosis and cell senescence while S1P induces cell migration, survival, proliferation, angiogenesis and inflammation [6-9]. Therefore, SphKs act as a rheostat of ceramide, sphingosine and S1P; SphKs play a crucial role in determining the fate of a cell.

These sphingolipid metabolites with opposing biological effects are inter-convertible and their balance is modulated through the actions of SphKs. SphKs exists in two isoforms, SphK1 and SphK2. SphK1 has been shown to be overexpressed in many human tumors, including breast cancer, where it contributes to malignant progression [6,32,33]. SphK1 is predominantly located in the cytosol and can be stimulated by various growth factors and cytokines [7,31,34]. When SphK1 is activated through phosphorylation at residue Ser225 by ERK, it is translocated to the plasma membrane where it preferentially acts on its substrate, sphingosine [7,31]. S1P generated by this process at the inner leaflet can function intracellularly as a second messenger or it can be exported out of the cell through ABCC1 transporter and then bind to S1PRs with high affinity in an autocrine and/or paracrine manner to promote proliferation, migration and angiogenesis [7,10,11,35,36]. Contradictory to the functions possessed by SphK1,
early studies showed that SphK2 induces apoptosis via its putative BH3 domain and suppresses cell proliferation [37]. Localization also differs between SphK1 and SphK2. While SphK1 is predominantly localized in the cytosol, SphK2 exists in the nucleus, endoplasmic reticulum, and mitochondria [37-39]. In addition, SIP generated by SphK2 acts independently from the S1PRs [37]. However, recent study demonstrated SIP produced by SphK2 regulates activity of histone deacetylases 1 and 2 (HDAC 1 and HDAC2), leading to increased transcription of specific genes [40]. Furthermore, SphK2 downregulation with siRNA was shown to inhibit proliferation and migration in tumor cells [41]. Therefore, although the mechanisms in which SphKs affect downstream signaling differs between the two isoforms, this evidence suggests both SphK1 and SphK2 are implicated in progression of the cancer.

S1PRs (S1P<sub>i</sub>) are members of G-protein coupled receptor (GPCR) family that mediates SIP effects on biological functions [7]. There are five known S1PRs and they have distinct cellular and tissue distribution as well as affinities toward SIP [42,43]. Each S1PR is coupled to specific G-proteins that regulate numerous downstream signaling pathways, making this pathway more complex and unique [44]. S1P<sub>1</sub> is known to couple with G<sub>i</sub>, which leads to the activation of phospholipase C (PLC), Ras, ERK, phosphoinositide 3-kinase (PI3K) and AKT and inhibition of adenylate cyclase (AC). S1P<sub>1</sub> and S1P<sub>3</sub>, both coupled with G<sub>i</sub>, G<sub>q</sub>, and G<sub>i</sub>, but their effects are contradictory. Binding of SIP to S1P<sub>1</sub> activates ERK (G<sub>i</sub>), PLC (G<sub>q</sub>) and Rho (G<sub>12/13</sub>) but not Rac (G<sub>i</sub>). Coupling of SIP<sub>2</sub> with SIP results in PLC (G<sub>i</sub>), Rac (G<sub>i</sub>) and Rac (G<sub>q</sub>), and SIP, are dependent on G<sub>i</sub> where it increases cell proliferation via Ras/ERK pathway and migration via PI3K/Rac pathways and reduces apoptosis by activating the AKT pathway [45-47].

Coupling of S1P<sub>1</sub> with G<sub>12/13</sub> inhibits cell proliferation, growth and migration, which migration is known to be dependent on activation of Rho and Rac-GAP [47]. S1P<sub>3</sub> is known to couple with G<sub>i</sub>, G<sub>q</sub>, and G<sub>i</sub>, where binding to G<sub>i</sub> activate Cdc42, PLC and ERK and binding to G<sub>q</sub>, results in Rho activation and inhibition of proliferation [47-50]. Although the understanding of the mechanisms is limited, S1P<sub>3</sub> has been demonstrated to couple with G<sub>i</sub> and G<sub>q</sub> [51]. Interestingly, S1P<sub>3</sub> has dual functions that are mediated through different pathways depending on the developmental stage of the cells. In brain, S1P<sub>3</sub> induces process retraction of premature oligodendrocyte while inducing cell survival in mature oligodendrocyte through G<sub>i</sub> and AKT activation [52]. Together, these observations illustrate various aspects of sphingolipid pathway including the sphingolipid rheostat and associated enzymes, and that S1PRs play important roles in determining cell fate.

Sphingolipid Signaling and Breast Cancer

The role of sphingolipid metabolism in breast cancer has recently been gaining interest. Infact, there is substantial evidence of a role for sphingolipid metabolites in numerous cancers, including breast cancers. For example, breast tumor biopsies from patients are reported to have a significantly higher expression of SphK1 than adjacent normal mammary epithelium [33]. In addition, the level of intracellular glucosylceramide was higher in drug resistant breast tumor patients [53]. Interestingly, Ruckhaberle et al. demonstrated that enzymes associated with ceramide/sphingosine and SIP are differentially expressed between ER-positive and ER-negative breast cancers. In this study, SphK1, ceramide galactosyltransferase, and ganglioside GD3-synthase displayed higher expression among ER-negative tumors while ER-positive tumors expressed higher levels of glucosylceramide synthase, dihydroceramide synthase and acid ceramidase [54]. These findings suggest the possible and viable target may differ between breast cancer subtypes. In the following sections, we summarize the role sphingolipid signaling in each of the four subtypes of breast cancer (Table 1).

Luminal A: ER/PR-positive and HER2-negative

SphK1 expression is well known to associate with estradiol-dependent mitogenic and carcinogenic action in human breast cancer [55,56]. Highly expressed SphK1 in primary ER-positive breast tumors is correlated with reduced breast cancer-specific patient survival and increased tamoxifen resistance [57]. Additionally, high membrane SIP expression in the breast tumor is associated with increased resistance to tamoxifen; high cytoplasmic SIP, and SIP expression is associated with tamoxifen resistance as well as reduced disease-specific survival. This evidence suggests that SphK1/SIP pathway is involved in the progression of the ER-positive breast cancer.

In vitro studies have demonstrated that overexpression of SphK1 in ER-positive and HER2-negative breast cancer MCF-7 cells have increased resistance to doxorubicin, tamoxifen and tumor necrosis factor (TNFα) [55,58]. Also, exogenous SIP has been shown to induce ERK activation and migration in MCF-7 [59]. Sphingolipid signaling in MCF-7 cells have been extensively studied to fill the black box between hormone stimulus (often estradiol) and its effect on tumorigenes due high ER expressions in these cells. Estradiol, a steroid hormone widely used to stimulate ER, activates SphK1 and increases both intracellular and extracellular SIP [60]. Released SIP is capable of binding to S1PRs as discussed earlier and activate downstream signaling in an autocrine and/or paracrine manner [11].

The most predominantly expressed S1PR in MCF-7 cells is SIP<sub>1</sub> and it is capable of being activated by both Estradiol and SIP [57,60-63]. Activated SIP<sub>1</sub> in turn, enhances downstream signaling including EGFR transactivation and ERK phosphorylation [57,60]. A recent study demonstrated that both Estradiol and SIP induce rapid internalization of membrane EGFR while sustaining cytosolic/ endosome EGFR levels, indicating a delay in degradation of the EGFR [43]. An important player in maintaining high endosomal EGFR level and inhibiting EGFR degradation is Cdc42. Overexpression of Cdc42 leads to metastasis and overall migration potential of breast cancer cells [64,65]. SIP interaction with SIP<sub>1</sub> induces Cdc42 activation, which inhibits binding of c-Cbl (E3 ubiquitin-protein ligase) to EGFR and consequently prevents c-Cbl from catalyzing EGFR ubiquitination and degradation. Compared to surface EGFR, early endosome localized EGFR promotes more sustained activation of the Ras/ERK pathway that is crucial for cell proliferation and activation.

Table 1: Summary of different sphingosine kinases/spingosine-1-phosphate (S1P)/S1P receptors functions in different subtypes of breast cancer.

<table>
<thead>
<tr>
<th>Luminal A (ER+/HER2--)</th>
<th>Luminal B (ER+/HER2+)</th>
<th>Triple negative (ER--/HER2--)</th>
<th>HER2 (ER--/HER2+)</th>
</tr>
</thead>
<tbody>
<tr>
<td>↑SphK1</td>
<td>↑SphK1</td>
<td>↑SphK1/2</td>
<td>↑SIP</td>
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<td>↓SIP</td>
<td>↓SIP</td>
<td>↓cycle</td>
<td>↓SIP/P</td>
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of ERK [66]. Phosphorylated ERK accumulates into membrane ruffles/lamellipodia and the nucleus and promotes migration of the cell. In the membrane ruffles, activated ERK promotes migration through the regulation of the actomyosin contractility while nuclear phosphorylated ERK enables cell movement by inducing metalloproteinase genes that are responsible for proteolytic degradation of the cell matrix [57,67]. Together, these studies suggest SphK1 activation by Estradiol induces S1P release and subsequent binding to S1P3, which sustains cytosomal EGFR and ERK activities to promote MCF-7 cell migration.

Luminal B: ER/PR-positive and HER2-negative

In contrast to ER-positive and HER2-negative breast cancer patients where high SphK1 expression has a negative impact on prognosis and resistance to chemotherapy, high SphK1 expression is correlated with increased patient survival and reduced recurrence in ER- positive and HER2-positive breast cancer patients being treated with tamoxifen [68].

Similar to the clinical evidence, ER-positive MCF-7 cells transfected with HER2 have been shown to tolerate oncogenic characteristics [57]. In these cells, SphK1 expression is increased in a HER-2 dependent manner compared to MCF-7 cells with control vector [57]. SphK1 in turn leads to reduction in HER2 expression and limits p21-activated protein kinase1 (p65 PAK1, downstream of HER2 and upstream regulator of ERK) and ERK expressions, both of which are known to induce migratory phenotype upon S1P treatment [59,69]. Upon S1P stimulus, HER2-overexpressed MCF-7 cells hold the capability of phosphorylating ERK in a S1P3-dependent mechanism [57]. However, unlike MCF-7 cells lacking HER2, phosphorylated ERK remains in the cytoplasm and the nuclear translocation or accumulation in the membrane ruffles/lamellipodia that leads to migration does not occur. These findings suggest that targeting HER2 and S1P3 in combination may represent a potential therapeutic avenue for ER-positive breast cancers. To date, emphasis has been placed on S1P1 and other S1PRs are not studied in details in HER2- and ER-positive breast cancers. Measuring S1PR expression and understanding mechanisms involving other S1PRs may elucidate the role of SphK1/S1P pathway in this breast cancer subtype.

Triple negative/basal like: ER-and HER2 negative

Compared with ER-positive breast cancers, ER-negative breast cancer patients have earlier disease recurrence and reduced survival times primarily due to the lack of response to hormonal therapies such as tamoxifen. Additionally, patients who are also negative for HER2 have more limited therapy selections because of insensitivity to immunotherapy such as Herceptin. Therefore, it is especially critical to identify novel targets for therapeutic intervention that can provide better treatment options for triple negative breast cancers.

SphK1 has been demonstrated to have a significant positive correlation with loss of ER expression and greater tumor aggressiveness and poorer prognosis in breast cancer patients [54]. Similarly, SphK1 is highly expressed in metastatic triple negative MDA-MB-231 breast cancer cells and its inhibition decreases cell cycle disruption and proliferation, and increases apoptosis [17]. This was evidenced in a study where downregulation of SphK1 increased intracellular sphingosine and its accumulation in the cytosol, which in turn inhibited cell cycle regulators, cdc2 activity and Chk1 [70]. Chk1 compromises spindle checkpoint function and cytokinesis [70]. Interestingly, other studies have shown stronger anticancer effects, including decreased cell viability, cell proliferation, migration/invansion, and induced apoptosis, by SphK2 ablation [17,41]. This is unexpected because downregulation of SphK2 with siRNA increases intracellular S1P level in MDA-MB-231 cells, most likely due to SphK1 compensation, which we assume to promote cell growth [41]. Furthermore, inhibition of SphK2 reduced pro-survival transcription factor, nuclear factor kappa-light-chain-enhancer of activated B cells (NFκB), through reduced activation of the Ser536 phosphorylation site on the p65 subunit, suggesting cross-talk between SphK pathway and NFκB might play a role in resistant characteristics [17].

These findings indicate both SphK1 and SphK2 are involved in ER-and HER2-negative breast cancer progressions, however, which isoform plays more significant role in tumor growth is yet to be elucidated. A change in either SphK1 or SphK2 expression can differentially alter other sphingolipid levels and cause either pro- or anti-cancer behaviors at downstream. However, roles of sphingolipid signaling in triple negative breast cancers are not well studied compared to ER-positive cancers, therefore whether S1PRs are involved and, which receptors have effects on tumor progression in triple negative breast cancer remains unclear.

HER2 type: ER-negative and HER2-positive

SphK1 also has a strong correlation with poor prognosis in ER-negative and HER2-positive breast cancers [18]. In a study with cohort of 140 ER-negative breast cancer patients, high expression of SphK1 in the HER2-positive tumors was associated with shorter disease- specific survival and disease-free survival compared to patients with low SphK1 expression in their HER2-positive tumors. Additionally, a high level of SphK1 and S1P3 are linked with shorter disease-free survival and disease-specific survival in ER-negative breast cancer patients.

In ER-negative and HER2-positive MDA-MB-453 cells, EGF has been demonstrated to phosphorylate both SphKs in an ERK dependent manner, leading to an increase in cell migration [71]. These cells express abundant S1P3, small quantities of S1P3/4 and very limited S1P4. Despite predominant expression of S1P3, the less abundant S1P1 interact with HER2 to regulate S1P-induced ERK [69]. This was evidenced by reduced S1P-stimulated ERK activity through independent downregulation of S1P3 and HER2 expressions. In addition, inhibition of SphK1 reduced S1P/S1P4-induced activation of ERK1/2 and altered HER2 trafficking in these cells [18]. Furthermore, inhibition of SphK1 reduced and treatment with a S1P1 agonist, phyto-S1P, stimulated ERK activation via a mechanism that involves HER2, suggesting synergistic interactions between S1P1 and HER2 in these cells [18,69].

In contrast to the findings in ER-positive cells where HER2 and SphK1 interact in a negative feedback mechanism to induce tolerance against cancer progression, the interactions of SphK1, S1P, and S1P4 with HER2 suggest sphingolipids act together with HER2 to enhance ER-negative breast cancer progressions. These findings indicate S1P3 and HER2 combination treatment is promising candidate for patients with ER-negative and HER2-positive characteristics. We can also suggest that the relative proportion, not the absolute quantity of the S1PRs may play critical roles in determining cell fate at least in this subtype as non-predominant S1P3 plays a critical role in ERK phosphorylation, which is a major factor affecting cell proliferation.
Conclusions

There is considerable information concerning the role of sphingolipid signaling in cell death and tumorigenesis in breast cancer. Interestingly, each subtype of breast cancer responds differently to the sphingolipid signaling cascade and thus possesses different effects on cell fate as well as resistance to chemotherapy. Specifically, previous studies have placed focus on S1P3 in ER-positive breast cancers although SphK1 is expressed more intensively in ER-negative/HER2-positive breast cancers and whether other S1PRs are involved remains unclear.

Moreover, two ER-positive subtypes, one positive and other negative for HER2, possess opposite effects on cell fate although S1P and S1P4 are induced in both subtypes. Additionally, fewer studies have placed emphasis on the role of sphingolipids in ER-negative breast cancers although SphK1 is expressed more intensively in ER-negative breast cancers. Further studies on sphingolipid signaling, especially involving SphKs, S1P and S1PRs, which are shown to cause unique biological effects in different subtypes, will help elucidate the mechanisms involved in tumorigenesis. Additional work will provide necessary information for precise and detailed prognosis and give more accurate chemotherapy selections for breast cancer patients in the future.

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