

Analysis of Autosomal SNPs in Paraffin-embedded Tissue Samples: Two Cases

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Abstract

Despite the numerous STR loci available for analysis of cases submitted to forensic laboratories, namely with the use of commercial kits that amplify mini - short tandem repeats (STRs), some cases remain "unsolved" i.e. fail to yield a STR DNA profiling result or result in likelihood ratio calculations that are not sufficiently high. Autosomal single nucleotide polymorphisms (SNPs) may constitute a potential advantage to solve some of these difficult cases, as has already been described by others.

In this work we present two revisited cases of our routine work, where the type of samples available for analysis was histological blocks of paraffin-embedded tissue samples from a man (Case 1-"lymph node") and from a woman (Case 2). The application of autosomal SNPs in the analysis of paraffin-embedded tissues constitutes one of the possible approaches that can be used in our laboratory in some "difficult" cases, and is part of a validation process of the described methodology.

Following extraction the two sample DNAs had already been genotyped with the routine commercial STR or mini-STR kits, like AmpFLSTR® Identifier®, MiniFiler™ and NGM™ PCR Amplification Kits and compared to their respective reference samples. These samples were then typed for the 52 loci included in the SNP for ID 52 plex using a SNaPshot™ assay previously described by Sanchez et al. in order to test and validate this multiplex in real casework [1].

The study showed that autosomal SNP can be used as additional markers to the profiling of difficult samples such as paraffin embedded tissue.

Introduction

Currently forensic laboratories solve almost all the requested cases through the traditional use of commercial kits that amplify short tandem repeats (STRs) or mini-STRs. In spite of this, there are still some "unsolved" or challenging cases that are occasionally encountered in forensic laboratories like paraffin-embedded tissue. "Unsolved" or challenging cases can be paternity cases, usually where the father is deceased and other relatives are disposable to investigate the paternity, that often lead to low paternity indexes after STR profiling. Paraffin embedded samples are not usual in forensic investigations. But these types of samples due to their high level of degraded sample DNAs, may also lead to low Likelihood Ratios (LR) after conventional STR analysis.

The use of autosomal SNPs for forensic research has been widely discussed in recent years. SNPs have important advantages compared to short tandem repeats (STRs) [2] and in combination with standard markers, improve the power of discrimination and provide potential supplementary data for paternity testing [3,4]. Moreover their study may constitute an improvement in the analysis of degraded samples [5].



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M Lurdes Pontes^{1,2*}, Olalla Maroñas Amigo³,
Manuel Fondevila³, M. João Porto^{1,2} and R.
Medeiros^{4,5,6}

¹Instituto Nacional de Medicina Legal e Ciências Forenses, Portugal

²CENCIFOR- Forensic Science Center, Portugal

³Institute of Legal Medicine, Universidade de Santiago de Compostela, Santiago de Compostela, Spain

⁴Molecular Oncology group, Portuguese Institute of Oncology, Portugal

⁵ICBAS, Abel Salazar Institute for the Biomedical Sciences, University of Porto, Largo Professor Abel Salazar, 4099-003, Porto, Portugal

⁶LPCC, Research Department-Portuguese League Against Cancer (NRNorte), Estrada Interior da Circunvalação 6657, 4200 Porto, Portugal

Address for Correspondence

M Lurdes Pontes, Instituto Nacional de Medicina Legal e Ciências Forenses, Portugal, E-mail: mlurdes.pontesrebelo@gmail.com

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As it was already said, paraffin embedded samples are not usual in forensic investigations. Here, the only two interesting cases involving paraffin-embedded tissue samples we have had over the years are presented. These cases were previously resolved using the traditional STR markers, but they were revisited in order to access the relevance of studying autosomal SNPs. This was achieved by calculating the LR before and after adding the information of the analyzed SNPs. The use of SNPs for challenging forensic sample type's is documented by others, as is the recovery and analysis of DNA from formalin-fixed paraffin-embedded (FFPE) tissues but mostly in medical context and using other analysis methodologies. Equally, the combining of statistical data from STR and SNP loci has been demonstrated before.

In spite of these two cases the STR results alone would have provided sufficient certainty of source identification to be used without the additional SNP work, our aim was to validate the use of the SNP for ID 52 plex and demonstrate the increment of the calculated LRs after their use as additional markers to the profiling of difficult samples such as paraffin embedded tissue, in order to apply their use in similar cases that may be encountered in our forensic laboratory in the future.

Materials and Methods

Samples

In this study histological material from two persons with a histological tumor - positive diagnostic was available. The aim of the requested examinations was to confirm the identity of the persons involved, supposedly the donors of the paraffin-embedded samples,

who questioned the results of the histology laboratory biopsies observations. For that, a DNA profile had to be obtained from cuts of the histological blocks of paraffin-embedded tissue samples (Figure 1); then it was necessary to compare the obtained profiles with the ones of the reference samples from the donors of the histological samples, respectively a man and a woman.

Extraction

DNA of reference samples (buccal swabs) was extracted using the slightly modified Chelex® method [6] and the DNA from the casework samples was extracted using the EZ1 DNA Tissue Kit together with the EZ1 DNA Forensic Card protocols with a BioRobot® EZ1 (Qiagen) [7].

Quantification

DNA concentrations were determined by real-time PCR using the Quantifiler™ Human DNA Quantification Kit (AB) and the ABI PRISM® 7500 Sequence Detection System (AB). The final concentrations of the casework samples were 0.02 and 0.05 ng/µl, respectively.

PCR amplification

The STR and mini-STR amplification of the extracted samples was achieved following the manufacturer protocols for the kits Identifiler® and Minifiler™ or NGM™ respectively.

A total of 52 autosomal SNPs were amplified by polymerase chain reaction (PCR) in 12.5 µl reactions containing DNA, in the case of these particular casework samples we adjusted the final volume to 25 µl in order to have a higher input of DNA sample. The PCR conditions were performed as previously described [1]. When using the problem samples of cases 1 and 2, we used a DNA input of ~0.10 and ~0.25 ng respectively.

SBE reaction

Two single base extension (SBE) reactions were performed (Auto 1 and Auto 2) as previously described with slight modifications [1] and included 2.5 µl SNaPshot reaction mix (Applied Biosystems - AB), 1.5 µl SBE primer mix (0.01–0.27 µM) and 2 µl of purified PCR product (with Exonuclease I/Shrimp Alkaline Phosphatase - ExoI-

SAP), totaling 6 µl. Excess nucleotides were then removed by the addition of 1 µl SAP (1 U/µl) to the SBE mix. The multiplexes were balanced in the laboratory in order to validate the described method in our particular conditions.

Typing

STR and SBE products separation was performed by capillary electrophoresis, using an ABI Prism 3130 xl Genetic Analyzer (AB), with 36 cm capillary arrays and POP-4 polymer (Applied Biosystems, Foster City, CA, USA). Analysis was made using GeneMapper® ID-X. For the autosomal SNPs allele calls were made manually.

Quality control

The 52 plex SNP assay was validated, locally and through inter-laboratory collaborative exercises, promoted by the GHEP-ISFG group.

Analysis of data

The LR's were calculated using an in house excel spread sheet, simply by multiplying the individual LR's from each of the SNP markers, using the previous published frequencies for those markers in a population sample of the North of Portugal, assuming that all the markers are independent [8].

Results

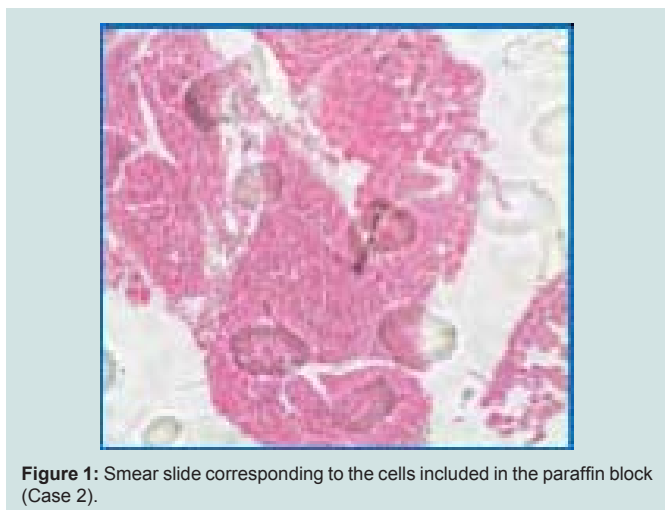
The results obtained for the STRs for Case 1 are presented in Table 1. These were only possible for 11 and 14 STR markers, respectively for case 1 and 2. Concerning SNPs, as it was expected, the results were possible for all the 52 typed SNPs in one of the cases (Case 1) and for 50 SNPs in the other (Case 2). The results for Case 2 are shown (Figure 2).

Table 1: STRs/mini-STRs results (Case 1).

STR marker	Paraffin embedded tissue cut	Reference Buccal swab
D8S1179	13	13
D3S1358	15,16	15,16
D19S433	14,16.2	14,16.2
D21S11	30,32.2	30,32.2
D7S820	11	11
CSF1PO	11,12	11,12
D13S317	12,13	12,13
D16S539	12	12
D2S1338	20,25	20,25
D18S51	12,18	12,18
FGA	24	24
Amelogenin	XY	XY

Table 2: Likelihood ratio calculation results.

Studied cases	Likelihood Ratio (LR)		
	STRs	SNPs	Total
Case 1	1.338E+15	9.046E+26	1.211E+42
Case 2	7.354E+19	4.682E+27	3.443E+47



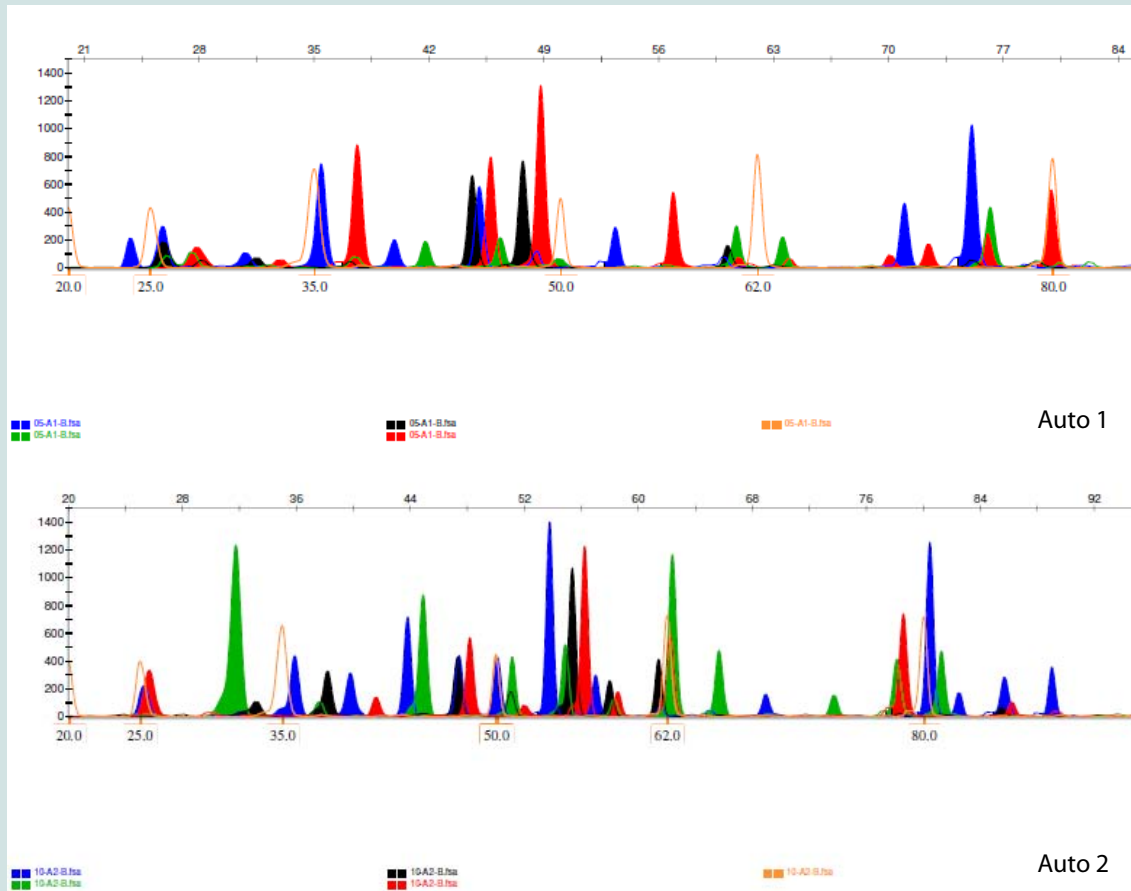


Figure 2: Electropherogram corresponding to the typing of the SBE products from the 52-plex Auto 1 and Auto 2 (Case 2).

The impact of studying SNPs in the overall Likelihood Ratio (LR) is presented by calculation of this parameter before and after adding the additional information given by the typed SNPs (Table 2), assuming all the studied loci are independent. Combining of statistical data from STR and SNP loci has been demonstrated before, but it is still not known if all SNP loci are independent of each other or of the STR loci used in this study. It is only known that this set of 52 SNP markers was carefully tailored in order to be sufficiently apart from each other guaranteeing this independence as reported by Sanchez et al. [1].

Before this example of autosomal SNP application some validation work was made in our laboratory, namely using internal controls of known standard DNAs and through inter-laboratory collaborative exercises, promoted by the GHEP-ISFG group. We also used allele frequencies for the 52-SNPs on our reference population – north of Portugal population [8].

Discussion and Conclusion

The novelty of this work consisted in the application of the 52-SNP multiplex previously described [1] for study paraffin-embedded tissue samples, traditionally not usual in forensic cases. In the medical field there are many studies that involve paraffin-embedded samples. Another study looked at paraffin-embedded tissue for identification

of the tissue source using a SNP multiplex, but with a more recent methodology [9].

The typing results and the performed LR calculations revealed that the 52 plex SNP assay may constitute a robust and sufficiently sensitive tool to complement the results from the traditional markers (STR and mini-STRs) in difficult forensic casework, which involves samples with material particularly degraded, like paraffin-embedded tissue samples, old cadaveric remains like bones as it was already reported [10], and other low-template DNA and highly degraded DNA from crime case samples involved in routine casework [11,12].

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